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Disclaimer



**General Colin Powell
Chairman (Ret), Joint Chiefs of Staff**



LESSON 4

A Leadership Primer

**"Don't be afraid to challenge the pros,
even in their own backyard."**

Learn from the pros, observe them, seek them out as mentors and partners.

But remember that even the pros may have leveled out in terms of their learning and skills.

Sometimes even the pros can become complacent and lazy.

Leadership does not emerge from blind obedience to anyone.

Xerox's Barry Rand was right on target when he warned his people that if you have a yes-man working for you, one of you is redundant.

Good leadership encourages everyone's evolution.

Preparing standards,
verifying analytical balance performance, and
pipetting
are fundamental lab techniques that are often assumed or simply overlooked.

Methods often "cookbook" how to prepare standards in very general terms but rarely provide the "hows" and "whys".

Methods also frequently ignore when is it appropriate to use volumetric pipettes, air-displacement pipettes or large-bore serological pipettes.

Objectives

Glassware Generalisms

Solution Solutions - Preparing Standards Perfectly

Proper Pipetting Principles

Balance Basics

Some Excellent Internet Resources

High School level pipetting

http://www.woodrow.org/teachers/esi/2002/Biology/Projects/lab_skills/ls7/

good close-up of micropipet tip when drawing and dispensing sample

http://www.rainin.com/pdf/edp_plus_manual.pdf

“Oliver’s Demos”

<http://www.csudh.edu/oliver/demos/pipetuse/pipetuse.htm>

<http://www.csudh.edu/oliver/demos/bal-use/bal-use.htm>

<http://acpcommunity.acp.edu/Facultystaff/genchem/GC1/lab/mvvolume/pipetech.htm>

<http://chemscape.santafe.cc.fl.us/chemscape/catofp/measarea/volume/pipet/pipet4.htm>

http://www.kimble-kontes.com/pdfs/class_a_b_tolerances.pdf

http://www.kimble-kontes.com/pdfs/reading_the_meniscus.pdf

http://www.kimble-kontes.com/pdfs/to_contain_to_deliver.pdf

<http://www.kimble-kontes.com/html/FAQ.html>

<http://www.kimble-kontes.com/html/RelatedLinks.html>

***** “Science By (Mr.) Jones”

<http://www.sciencebyjones.com>

***** “Chemistry Comes Alive!” (alphabetical topic search)

<http://jchemed.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/WORDS/WORDS16.HTM>

“Chemistry Comes Alive” sample

med.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/2QuantMenu/Transfer/TRANSOL/MENU.HTM

Transferring a 25-Milliliter Sample

A 25-milliliter volumetric pipet is used to remove a sample from a volumetric flask and transfer it to an Erlenmeyer flask.

Beginning of Quantitative Transfer with a Volumetric Pipet

 View Movie
4 MB
16 Seconds



 View Slides
4 Slide(s)

The task to be accomplished is explained.

Checking the Pipet

 View Movie
1.9 MB
67 Seconds



 View Slides
20 Slide(s)

A volumetric pipet is checked by filling it with deionized water and then observing as the water drains out.

Drain the Pipet Tip

 View Movie
8 MB
27 Seconds



 View Slides
10 Slide(s)

Water is removed from the tip of the clean pipet.

"Chemistry Comes Alive" sample

Rinse the Pipet with Solution

 View Movie
1.7 MB
61 Seconds



 View Slides
18 Slide(s)

The pipet is rinsed with the solution that is to be measured.

Fill the Pipet

 View Movie
2 MB
70 Seconds



 View Slides
17 Slide(s)

A pipet bulb is used to fill the pipet with the solution to be measured, and then the volume is adjusted to the mark using a finger to control the level of solution.

Transfer the Liquid

 View Movie
8 MB
29 Seconds



 View Slides
7 Slide(s)

The liquid is transferred to an Erlenmeyer flask by allowing it to drain naturally from the pipet.

Rinse the Last Drop

 View Movie
1.3 MB
44 Seconds



 View Slides
10 Slide(s)

The final drop of solution delivered by the pipet is rinsed into the flask.

CLASSWARE GENERALISMS

TC glassware

- TC = To Contain
- entire contents = the correct volume
- transferring the contents to another container requires a quantitative transfer.
- Example: Volumetric flask

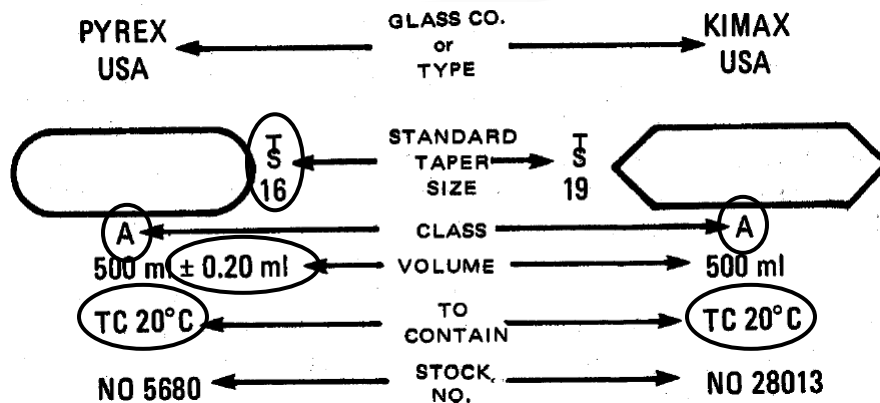
TD glassware

- TD = To Deliver
- **Generally DO NOT blow out the small amount remaining in the tip**
- **A flowtime must be observed.**
 - Allows all of the water film on the inside of the pipet to drain off, so you get the full accuracy the pipet is capable of.
 - Really accurate pipets, like class A, designed to drain so slowly that the film draining keeps up with the bulk draining.

Flowtimes for TD pipets

Nominal Volume mL	Class A	Class B
	Flowtime (sec)	Flowtime (sec)
1	10	3
10	15	8
25	25	15
50	25	15
100	30	30

Glassware labeling

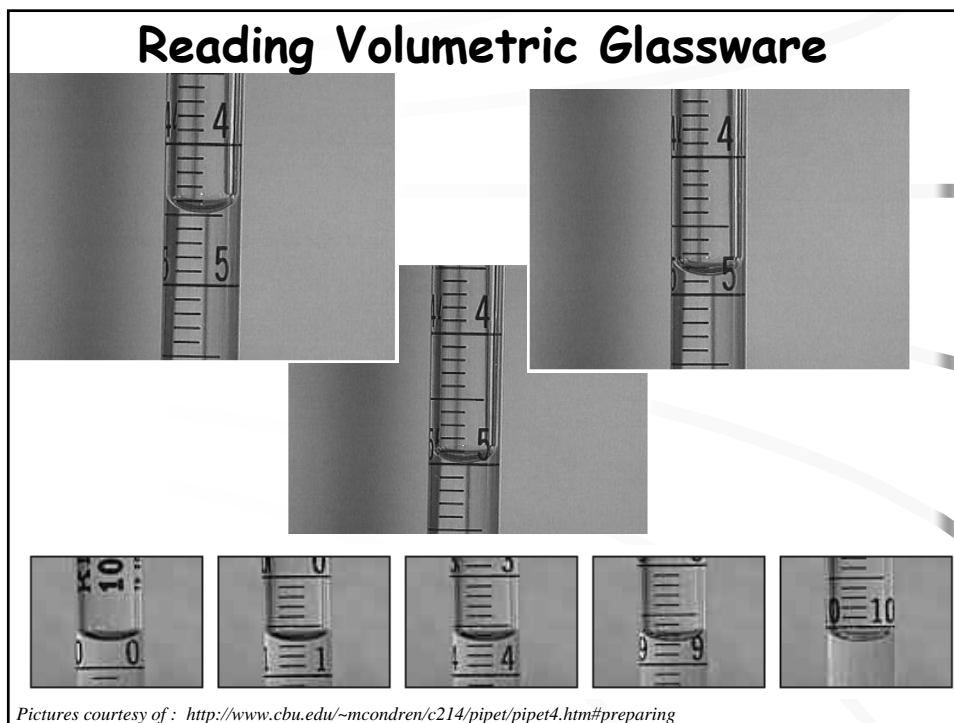
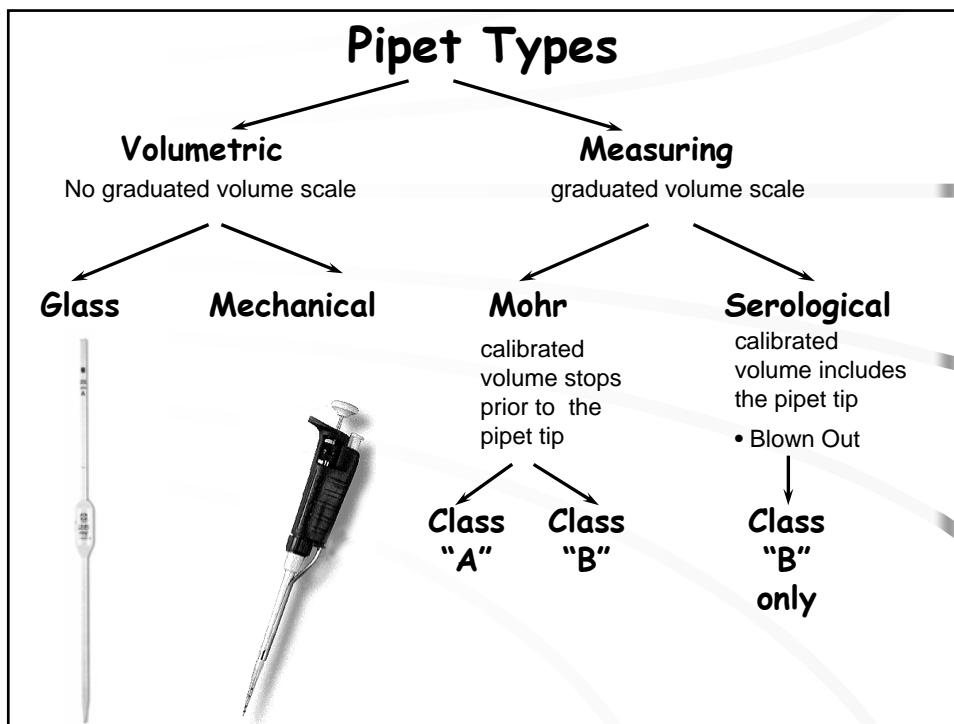


T = Standard Taper. Finished to a 1:10 taper. A single number following indicates the approximate diameter (mm) of the neck.

§ = Product Standard. Used for stopcocks with Teflon plugs. Finished to a 1:5 taper. A single number following indicates the diameter (mm) of the hole in the plug.

Glassware accuracy

Volumetric Flask Tolerances (mL)			Pipet Tolerances (mL)			Graduated Cylinder Tolerances (mL)		
Nominal Volume mL	Class A tolerance	Class B tolerance	Nominal Volume mL	Class A tolerance	Class B tolerance	Nominal Volume mL	Class A tolerance	Class B tolerance
10	± 0.02	± 0.04	1	± 0.006	± 0.012	10	± 0.08	± 0.1
25	± 0.03	± 0.06	5	± 0.01	± 0.02	25	± 0.14	± 0.3
50	± 0.05	± 0.10	10	± 0.02	± 0.04	50	± 0.20	± 0.4
100	± 0.08	± 0.16	25	± 0.03	± 0.06	100	± 0.35	± 0.6
250	± 0.12	± 0.24	50	± 0.05	± 0.10	250	± 0.65	± 1.4
500	± 0.20	± 0.40	100	± 0.08	± 0.16	500	± 1.10	± 2.6
1000	± 0.30	± 0.60				1000	± 2.00	± 5.0
2000	± 0.50	± 1.00				2000	----	± 10.0



What to use...and when

Glassware	Proper Use
Disposable beakers	For pouring out a small volume of standard to warm up and use.
Glass beakers or Erlenmeyer flasks	Digestions. NOT for standard preparation or anything requiring volumetric measurements.
Graduated Cylinders	BOD & TSS when using 100 mLs or more of sample
Micro-bore Volumetric pipets (glass or mechanical)	Standard preparation
Mohr pipets	Color reagent (phosphorus)- if using the NCL modification
Serological pipets (wide-bore)	BOD influents (low volume samples) also preservation of samples for phosphorus or ammonia
Wide bore volumetric pipets	BOD & TSS samples



$$C_1 V_1 = C_2 V_2$$

(Concentration of existing solution)

X

(Volume of existing solution)

=

(desired concentration of new solution)

X

(desired volume of new solution)

Solution Preparation Example 1

You have the following stock standards of phosphorus available:

50 ppm
5.0 ppm

You need to prepare: 50 mLs of a a 0.1 ppm standard

C_1 : Concentration of existing solution	50 ug/mL (or 5)		C_2 : Concentration of new solution	0.1 (ug/mL)
X	X	@	X	X
V_1 : Volume of existing solution	V_1 : unknown		V_2 : Volume of new solution	50 mL (need at least 50 mLs for a std)
	$50V_1 = 50 \times 0.1$			$5V_1 = 50 \times 0.1$
	$50V_1 = 5.0$			$5V_1 = 5.0$
	$\frac{50}{50} = \frac{5.0}{50}$			$\frac{5}{5} = \frac{5.0}{5}$
	$V_1 = 0.1 \text{ mLs}$			$V_1 = 1.0 \text{ mLs}$


Thus, 0.1 mLs of a 50 ppm standard, diluted to 50 mLs = 0.1 ppm

Standard Preparation Dilemma


		50. ug/mL	0	0.1	0.2	0.4	0.6	0.75	1.0 mL
0	50 mLs	25. ug/mL	0	0.2	0.4	0.8	1.2	1.5	2.0
0.1	final	5.0 ug/mL	0	1	2	4	6	7.5	10
0.2	volume								
0.4									
0.6									
0.8									
0.75									
1.0									
ug/mL									
	100 mLs	50. ug/mL	0	0.2	0.4	0.8	1.2	1.5	2.0 mL
	final	25. ug/mL	0	0.4	0.8	1.6	2.4	3.0	4
	volume	5.0 ug/mL	0	2	4	8	12	15	20

Which pipet(s) will I need?


Volume change of a cold solution-1




00.00



05.11



16.15



23.41

Volume change of a cold solution-2



$\frac{25 + 0.8 \text{ mLs}}{25 \text{ mL flask}} = + 3.2\%$



Quantitative Transfer Techniques

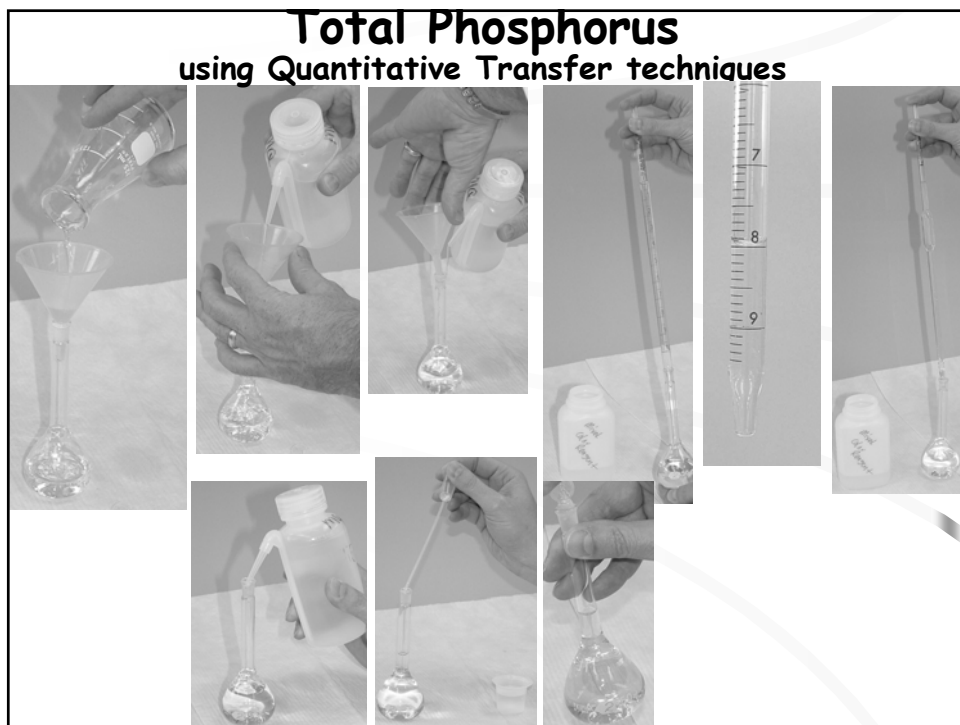


Be careful about the force of the stream...you do not want to cause particles to "fly" off the weighing dish
↓

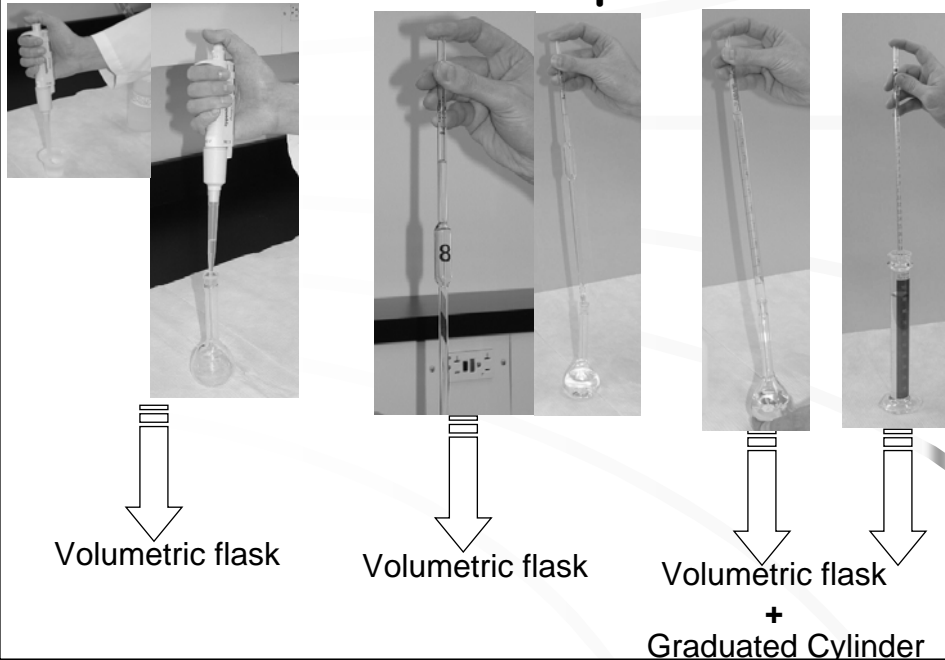


↑ Probably not a good technique for phosphorus due to potential for contamination...but a good general technique

Pictures courtesy of : <http://jchemed.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/1ChemLabMenu/Quantitative Transfer/>



Standard Curve Experiment



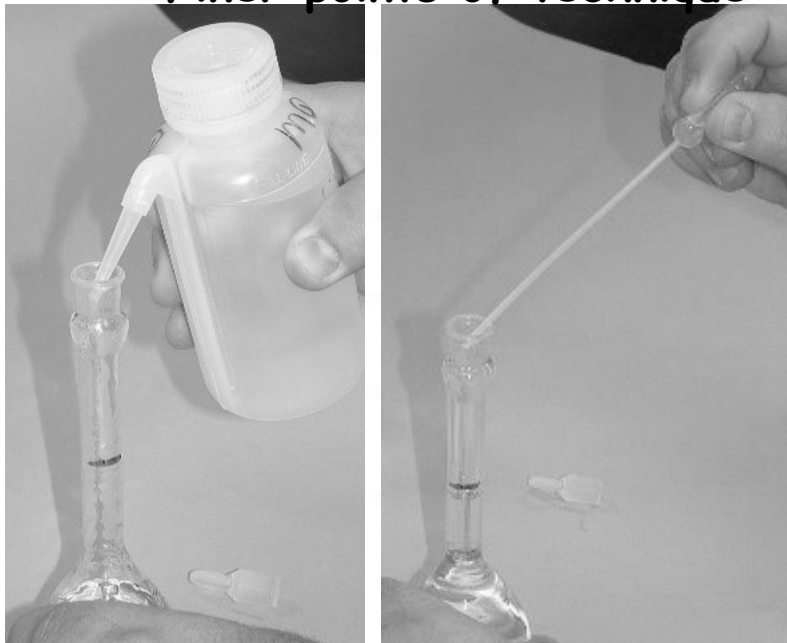
Standard mg/L P	Abs	Abs	Abs	Abs	Standard Curve Preparation Approaches
0	-0.001	0.001	0.001	0.007	
0.1	0.066	0.065	0.061	0.073	
0.2	0.132	0.129	0.114	0.135	
0.4	0.262	0.263	0.226	0.279	
0.5	0.328	0.329	0.282	0.354	
0.75	0.488	0.492	0.423	0.516	
1	0.652	0.652	0.55	0.687	
	<u>ADP/VF</u>	<u>VP/VF</u>	<u>MP/VF</u>	<u>MP/GC</u>	
	0.6515	0.6534	0.5508	0.6834	
	0.0007	0.0005	0.0046	0.0050	
	0.99999	0.99998	0.99984	0.99985	
	Air-displacement Pipet & 100 mL Volumetric Flasks				
	Volumetric Pipets with 100 mL Volumetric Flasks				
	Mohr Pipets with 100 mL Volumetric Flasks				
	Mohr Pipets with 100 mL graduated cylinders				

Standard Curve Preparation Approaches

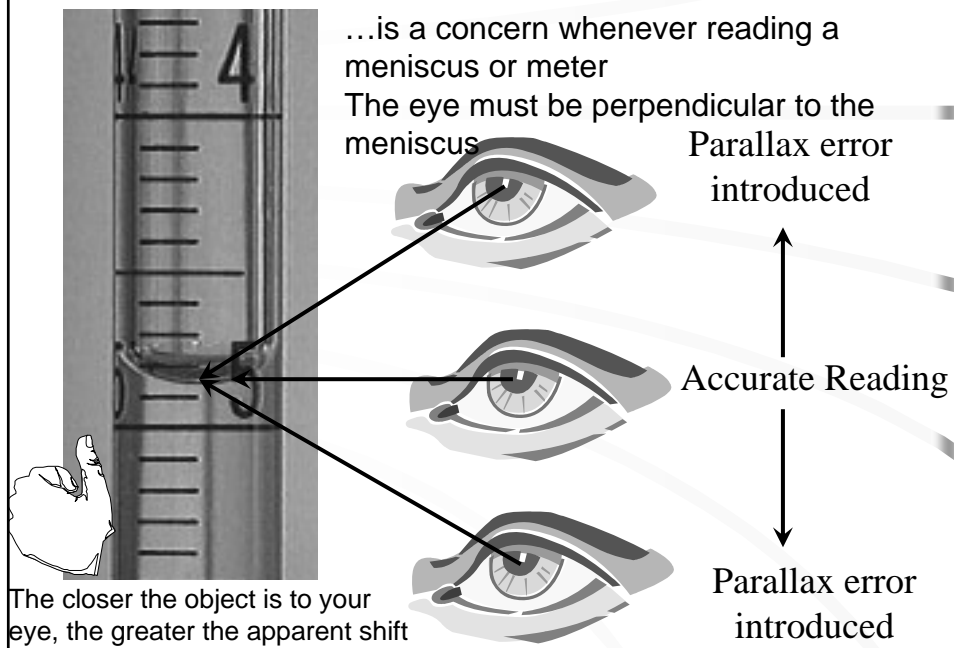
Air-displacement Pipet & 100 mL Volumetric Flasks
 Volumetric Pipets with 100 mL Volumetric Flasks
 Mohr Pipets with 100 mL Volumetric Flasks
 Mohr Pipets with 100 mL graduated cylinders

Standard mg/L P	RF	RF	RF	RF
0				
0.1	0.660	0.650	0.610	0.730
0.2	0.660	0.645	0.570	0.675
0.4	0.655	0.658	0.565	0.698
0.5	0.656	0.658	0.564	0.708
0.75	0.651	0.656	0.564	0.688
1	0.652	0.652	0.550	0.687
Range	0.651 - 0.660	0.645 - 0.658	0.550 - 0.610	0.675 - 0.730
mean RF	0.656	0.653	0.571	0.698
stdev	0.003912	0.005064	0.020472	0.019356
%RSD	0.60%	0.78%	3.59%	2.77%

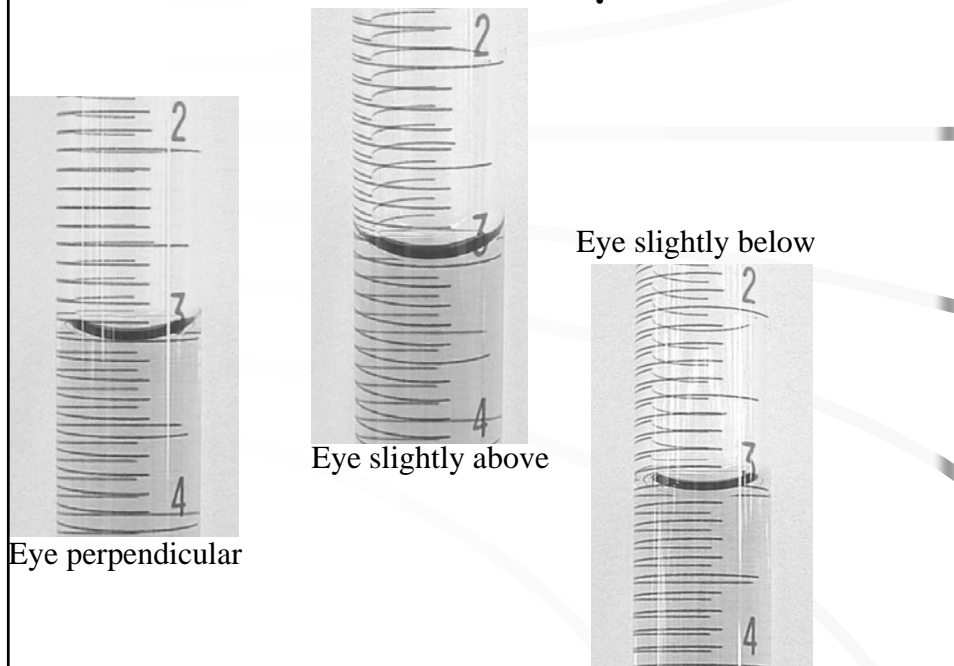
Finer points of technique



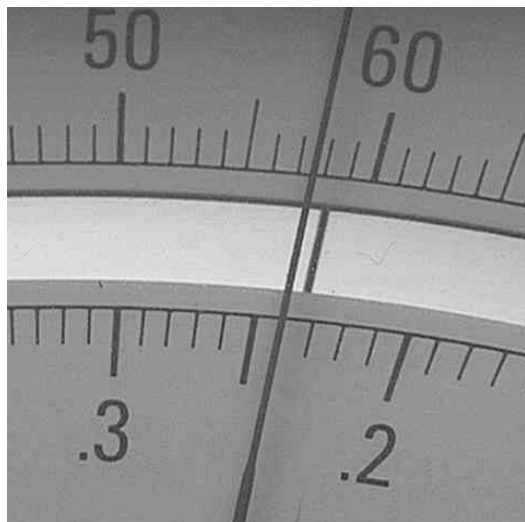
Parallax error...



Parallax error...up close



Parallax error and meters



Pictures courtesy of : http://jchemed.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/1ChemLabMenu/Measuring/Spectroscopy/spec20_menu/spec20_X_06362610/PICTURE.HTM?3

Value of disposable "beakers"



...a slight added cost to the laboratory, but an excellent way to minimize contamination and the time it takes to come to room temperature

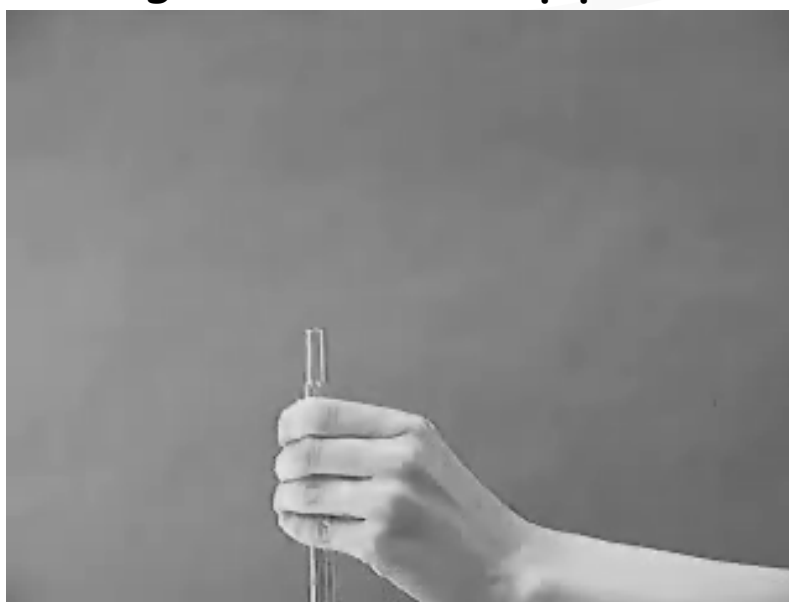
Solution Preparation Summary

- What equipment / standards do you have to accomplish the task?
- $C_1V_1 = C_2V_2$
- Bring standards/reagents to room temp. before use
- Never pipet from the reagent/standard bottle
- Ensure standards & reagents are properly labeled and linked to un-expired standards
- Read the meniscus properly
- Errors are additive

Pipettes & Pipetting

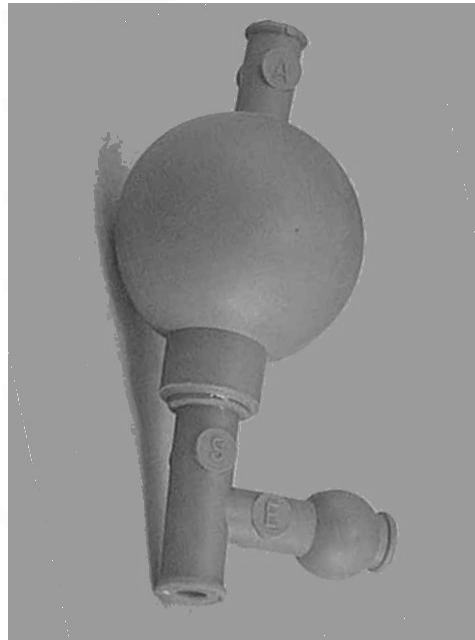
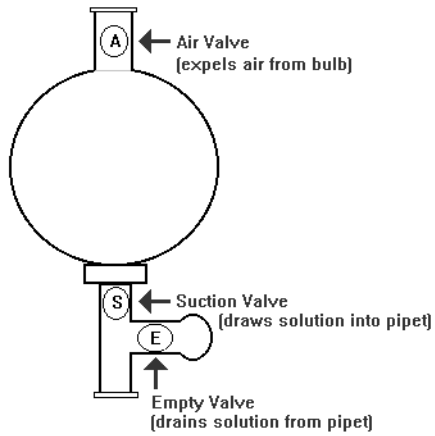


Using a conventional pipet bulb



Video courtesy of
<http://jchemed.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/1ChemLabMenu/Measuring/Volume/Pipet/PipetBulb/bulb.html#MENU.HTM>

Using a 3-Way bulb



Using a 3-Way bulb



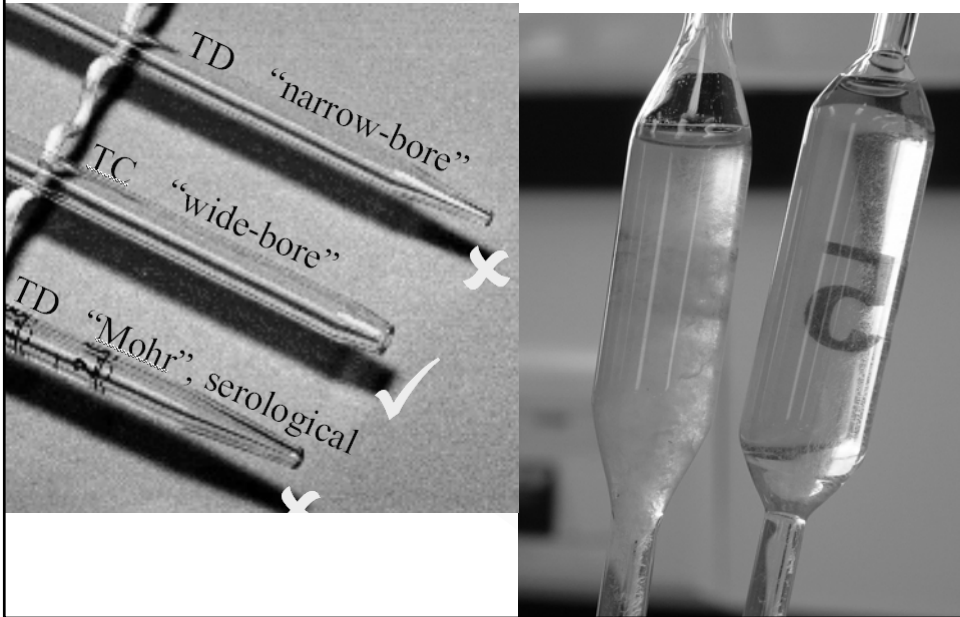
Video courtesy of

http://jchemed.chem.wisc.edu/JCESoft/CCA/CCA6/MAIN/1ChemLabMenu/Manipulating/TransferringSamples/Quantitative/TransferringLiquid/mohr_menu/mohr_X_PIPTHREEWAY/THUMBS.HTM

Bulb Alternatives



Widebore pipets



Proper use of volumetric glass pipets

Key points:

- Use class A pipets to prepare standards
- Volumetric pipets are calibrated to deliver (designated -TD) a specific volume
- The inside and outside of the pipet tip must be dry or rinsed with the solution to be transferred before use

Use of a volumetric pipet

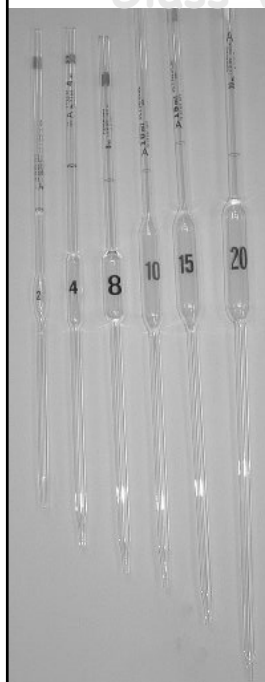
- Evacuate the pipet bulb by squeezing.
- Immerse the tip of the pipet into solution to be delivered
- Seat the bulb opening over the top opening of the pipet
- Hold the bulb in place while slowly releasing the squeezing pressure
- Continue to release the pressure while the solution is drawn into the pipet
- Draw the solution up well past the calibration line
- Curse when you go too far and draw the solution up into the bulb
- Quickly remove the bulb and seal the top of the pipet with the index finger
- Keeping the index finger in place, remove the tip from the solution
- Rest the tip of the pipet on the side of the container that held the solution.
- Slowly release finger; allow solution level (meniscus) to drop to calibration line.
- Place tip of the pipet over the receiving vessel and completely release the finger
- Keep the pipet upright and allow to drain completely (Note: Many class A pipets have the drain time imprinted adjacent to the "TD" designation.)

When the draining is complete, touch the tip of the pipet to the inside wall of the vessel and give it a half twist

Electronic v. mechanical pipet



Glass Volumetric vs. Adjustable volume mechanical



Using an autopipet

- 

1. SELECT THE CORRECT MICROPIPETTE FOR THE JOB. NOTE THE VOLUME RANGES ON THE PLUNGER.
- 

2. DIAL THE VOLUME.
- 

3. PLACE A TIP ON THE MICROPIPETTE WITHOUT TOUCHING THE TIP.
- 

4. DEPRESS THE PLUNGER TO THE FIRST STOP.
- 

5. PLACE THE TIP BELOW THE LIQUID.
- 

6. SLOWLY RELEASE THE PLUNGER.
- 

7. PLACE THE TIP NEAR THE BOTTOM OF THE EMPTY RECEIVING TUBE.
- 

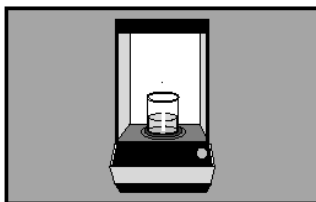
8. SLOWLY DEPRESS THE PLUNGER PAST THE FIRST STOP TO THE SECOND STOP.
- 

9. RELEASE THE PLUNGER AFTER THE TIP IS REMOVED FROM THE TUBE.
- 

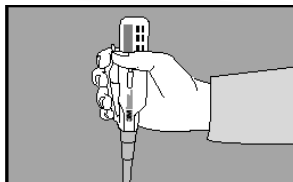
10. EJECT THE TIP INTO A WASTE CONTAINER.

Courtesy of: <http://www.thrcr.org/education/hutchlab/lessons/>

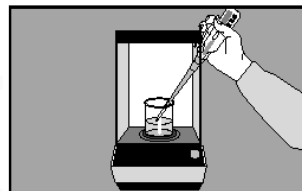
Checking autopipet accuracy



1. Place a disposable weighing dish that can hold 25-50 mLs on the balance. Prepare a clean disposable beaker containing reagent water at 20-25°C. Record the water temperature. Tare the balance to zero.



2. Aspirate a fixed volume of water from the vessel on the analytical balance. Note its weight. Tare the balance to zero.



3. Expel the water into the vessel on the analytical balance. Note its weight.
4. Repeat steps 2-3 for a total of 10 (or 20) measurements.
5. Calculate accuracy

Material condensed from: www.3m.com/microbiology/home/products/pipettor/calib.pdf

"Fast is fine... but accuracy is everything"

Wyatt Earp

Table 1: Artel's suggested initial tolerance limits

Pipette Volume ^(a)	Inaccuracy ^(b)	Imprecision ^(b)
2 µL	5.0%	2.0%
10 µL	2.5%	1.5%
20 µL	2.0%	1.0%
100 µL	1.6%	0.8%
200 µL	1.6%	0.8%
1000 µL	1.6%	0.8%

(a) Volumes are for variable volume pipettes at their maximum settings and for fixed volume pipettes. For variable volume pipettes, percentage error increases at lower settings; tolerance limits should be adjusted accordingly.

(b) These inaccuracy and imprecision values are double the specifications given by some of the major pipette manufacturers. Where manufacturers have differing specifications, the higher values were chosen.

Be prepared to
have auditors
inspect your
autopipets!

Information courtesy of: <http://www.artel-usa.com/Documents/Reports/report6.htm>

Pipet Summary

1. Use ONE pipet for the job (*error is additive*)
10 mL + 10 mL may get you 20,
but also gets you double the error
2. LESS volume is MORE chance for error
3. Use the proper pipet for the job
Mohr vs.
serological vs.
volumetric

Wide-mouth (BOD, TSS) v.
Microbore (standards, ammonia)
4. Maintain & calibrate your autopipettors

Balances

Balance Overview

Types of Balances

Selecting the right balance for the application

Care & Maintenance

Use

Preliminary Considerations

Making Absolute Measurements

Measurements and the Tare function

Select the right balance for the job

Accuracy Select a balance appropriate for the application.

Req'd

0.01 g Heavy objects (platinum crucibles, large flasks) -
Use a toploader type balance with at least 2 decimal
place (0.01 milligram) resolution and accuracy.

0.01 g Ascorbic acid reagent for Total Phosphorous -
Use a toploader type balance with at least 2 decimal
place (0.01 milligram) resolution and accuracy.

0.000,0 g TSS -
Use an analytical balance with at least 4 decimal
place (0.1 milligram) resolution and accuracy.

0.000,0 g Testing electronic or air-displacement pipettors -
Use an analytical balance with at least 4 decimal
place (0.1 milligram) resolution and accuracy.

Balance Use 1

Balances are not a “Plug n’ Play” device

1. If using an electronic balance, allow to warm up for at least 60 minutes.

Balance must be level to function properly

2. Check the balance leveling gauge to make sure bubble inside the target.



Balances need controlled temperature & humidity

Wide swings in humidity & temperature can damage the sensitive parts and electronics.

Balance Use 2

The pan and balance floor must be clean.

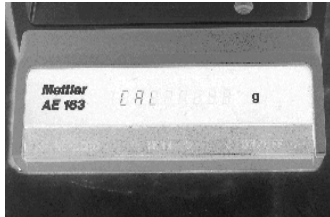
Use a camel-hair brush (keep near balance).

3. Dust off the balance pan with a clean soft -- *preferably camel-hair*-- brush. Use a mild detergent, DI water and lint free wipe if necessary.



Balance Use 3

4. Perform the internal calibration process if the balance has an on-board calibration function.



5. Zero the balance by pressing the "tare" bar (or button)



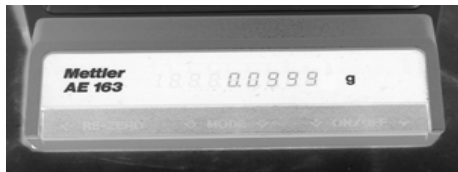
Balance Use 4

6. Place the first Class 1 weight on the clean balance pan ...



... with plastic forceps...

Balance Use 5



...allow the balance to stabilize

...measure and record the observed weight in the logbook.



Balance Use 6

7. Repeat step 6 with the other appropriate weights



8. Compare the observed weights to the acceptance ranges for the Class 1 weights. If any weight exceeds the acceptable range, **discontinue using the balance and take corrective action.**

Analytical Balance - Other Considerations

Sample must be at room temperature

warm or hot objects placed in a pan within a closed balance chamber can create air currents that buoy the pan, resulting in erroneous measurements

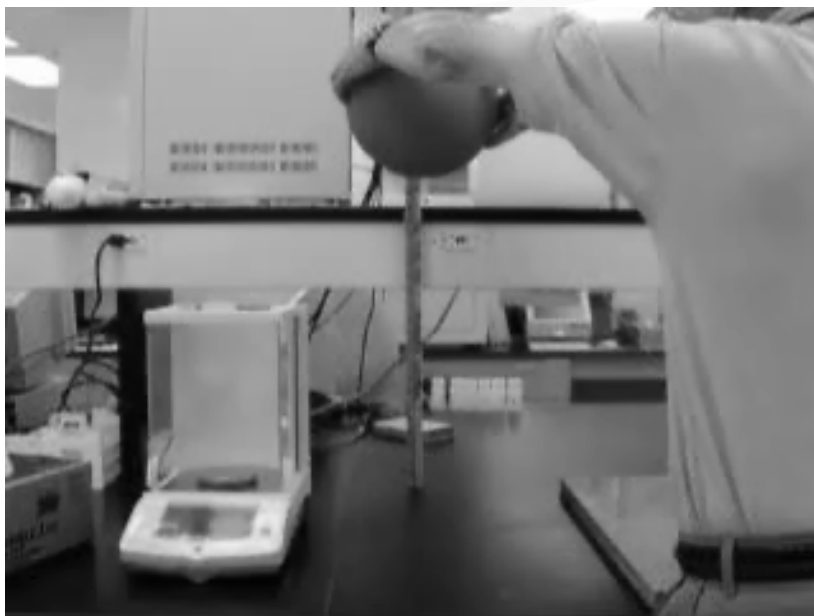
Sliding doors must be closed.

Avoid the effects of any draft or air currents on the balance

Protect the balance from vibration.

Do not bump the balance or table while making measurements. Vibration can significantly affect measurement accuracy...even leaning on a balance table without a damping pad can affect results by several milligrams.

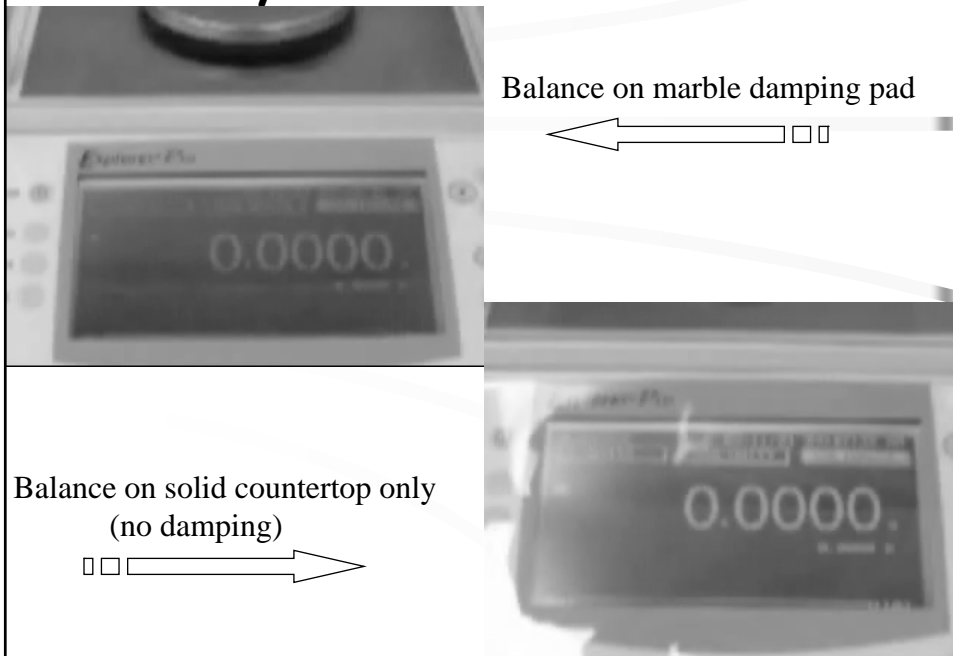
Vibration Test



Toploader Vibration



Analytical Balance Vibration



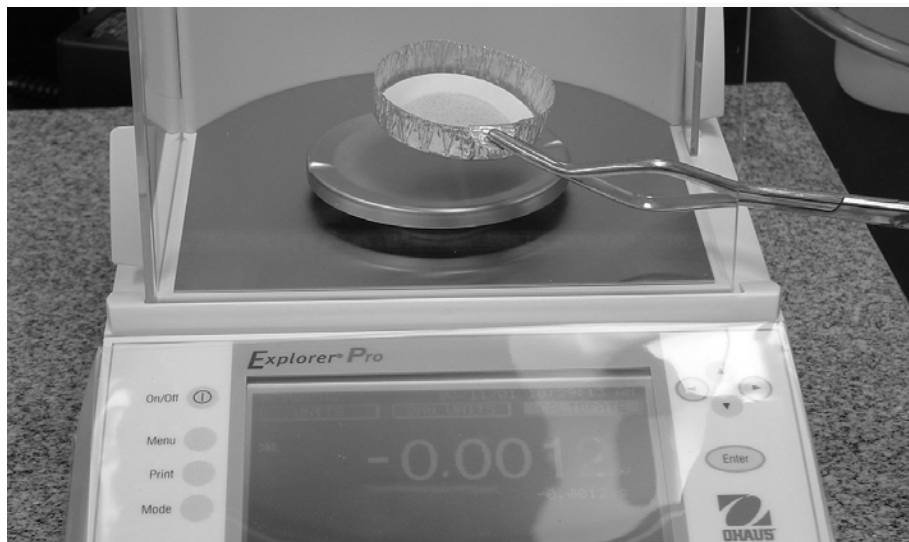
Balance on marble damping pad

Balance on solid countertop only
(no damping)

Analytical Balance Stability



Correct Procedure



*Determining Acceptance Ranges for Class 1 Weights
Using a Statistical Approach*

- Use the following guidelines until statistical limits have been established (*but check w/ your auditor first!*):
 - ▶ ± 0.3 mg* for weights <10mg
 - ▶ ± 0.5 mg for weights from 10mg to 100mg
 - ▶ $\pm 1-2\%$ for weights >100 mg
- Make 20 replicate measurements of the class 1 weight
- Spread measurements out over several non-consecutive days for best results
- Determine the mean and standard deviation (sd)
- The **upper acceptable range** = certified weight + 3 sd
- The **lower acceptable range** = certified weight - 3 sd

*Example 1
Determining the Acceptable Range for a 100 mg (0.1000 gm)
Class 1 Weight**

Item	Weight (grams)
Mean of 20 measurements	0.09999
Standard Deviation	0.000064
3 Standard Deviations	0.00019
Certified Weight	0.099998
Upper Acceptable Range	$0.099998 + 0.00019 = 0.1002$
Lower Acceptable Range	$0.099998 - 0.00019 = 0.0998$

Range = 0.0998 – 0.1002 g

***Balance readability of at least 0.0001 g**

Summary of Balance Accuracy Verification

- ☞ Calibrate balance daily if it is equipped with an on-board internal calibration feature
- ☞ Check balance accuracy monthly in the g and mg range
- ☞ Use certified class 1 (“S”) weights
- ☞ Have balance serviced annually
- ☞ Have weights re-certified annually or before expiration date
- ☞ Determine the acceptance range using a statistical approach or use the reasonable tolerance guidelines
- ☞ Always record weight measurements, whether they passed or failed and corrective action in the logbook
- ☞ **NEVER** use a balance that fails the verification check!!

Errors are additive

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Parallax errors

Cold vs. 20° C
Class A or not
Reading meniscus

Cold vs. 20° C
Class A or not
Reading meniscus

Cold vs. 20° C
Class A or not
TC vs. TD
1 volume → 2 pipets
macro v. microbore

Convective errors
Vibrational error
balance/weight accuracy

Quantitative transfers
multiple transfers

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General Colin Powell
Chairman (Ret), Joint Chiefs of Staff



LESSON 7

A Leadership Primer

"Keep looking below surface appearances.
Don't shrink from doing so (just) because you
might not like what you find."

"If it ain't broke, don't fix it" ...

...is the slogan of the complacent, the arrogant or the scared.

It's an excuse for inaction, a call to non-arms.

It's a mind-set that assumes (or hopes) that today's realities will continue tomorrow in a tidy, linear and predictable fashion.

Pure fantasy. In this sort of culture, you won't find people who pro-actively take steps to solve problems as they emerge.

Here's a little tip: don't invest in these companies.